# **CASE REPORT**

# **Open Access**

# Overexpression of plasmalemmal vesicle-associated protein-1 in patient with cyanotic nephropathy: a case report



Yusuke Ushio<sup>1\*</sup>, So Hirata<sup>1</sup>, Shun Manabe<sup>1</sup>, Mayuko Suyama<sup>1</sup>, Ayano Tanaka<sup>1</sup>, Momoko Seki<sup>1</sup>, Haruka Kato<sup>1</sup>, Kana Nomura<sup>1</sup>, Anna Nakai<sup>1</sup>, Hitoko Sumori<sup>1</sup>, Yuki Kawaguchi<sup>1</sup>, Shizuka Kobayashi<sup>1</sup>, Shiho Makabe<sup>1</sup>, Hiroshi Kataoka<sup>1</sup>, Naoko Itoh<sup>2</sup>, Sekiko Taneda<sup>2</sup>, Kazuho Honda<sup>3</sup> and Junichi Hoshino<sup>1</sup>

# Abstract

**Background** Cyanotic nephropathy (CN) is a known complication of cyanotic congenital heart disease (CCHD). However, many aspects of its pathophysiology remain unclear.

**Case presentation** We report the case of a 29-year-old male with a history of tetralogy of Fallot. Renal biopsy revealed glomerular hypertrophy and focal segmental glomerulosclerosis. Electron microscopy revealed extensive endothelial cell damage. To investigate the etiology of endothelial cell damage, PAL-E staining was conducted, revealing staining along the glomerular capillary wall.

**Conclusion** This is the first report of PAL-E staining in CN, suggesting potential overexpression of PV-1. The association of PV-1 expression with endothelial cell damage indicates its role in the pathogenesis of CN.

Keywords Cyanotic nephropathy, PAL-E, Cyanotic congenital heart disease, Case report

# Introduction

Cyanotic nephropathy (CN) was first documented in 1953 by Meessen and Litton, who reported 28 autopsy cases of cyanotic congenital heart disease (CCHD) and observed a correlation between glomerular hypertrophy and the degree of cyanosis [1]. CN has long been recognized as a potential complication of CCHD, with rising concerns about the increasing number of patients

\*Correspondence:

developing nephropathy as the survival rate of CCHD patients improves [2].

The pathogenesis of CN is hypothesized to involve increased shear stress from polycythemia, which induces nitric oxide (NO) release and subsequent glomerular hypertrophy. However, despite glomerular hypertrophy being the primary pathological finding, a previous study found no association between glomerular size and CN development when comparing CN and non-CN patients [3]. Autopsies have revealed pathological findings such as glomerular hypertrophy, capillary dilatation, an increased number of glomerular capillaries, thickening or rupture of capillary walls, and mesangial expansion [4–6]. However, a comprehensive study of the pathophysiology of CN has yet to be conducted.

Pathologische Anatomie Leiden-Endothelium (PAL-E) staining colors veins and peritubular capillaries but does not typically stain glomerular endothelial cells. There have been reports of PAL-E staining observed in



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

Yusuke Ushio

ushio.yusuke@twmu.ac.jp

<sup>&</sup>lt;sup>1</sup> Department of Nephrology, Tokyo Women'S Medical University, 8-1,

Kawada-Cho, Shinjuku-Ku, Tokyo, Japan <sup>2</sup> Department of Surgical Pathology, Tokyo Women'S Medical University,

<sup>8-1,</sup> Kawada-Cho, Shinjuku-Ku, , Tokyo, Japan

<sup>&</sup>lt;sup>3</sup> Department of Anatomy, Showa University School of Medicine, 1-5-8 Hatanodai, Shinagawa-ku, Tokyo, Japan

# **Case presentation**

A 29-year-old male with severe tetralogy of Fallot presented with 1+microscopic hematuria, which had worsened over the past year. Concurrently, urinary testing revealed proteinuria escalating from 1+to 3+during the same period. We received a consultation from a pediatric cardiologist; and laboratory tests showed a creatinine (Cre) level of 0.77 mg/mL; eGFR of 98.2 mL/min/1.73m<sup>2</sup>; blood urea nitrogen (BUN) of 16.2 mg/dL; 1+proteinuria; and microscopic hematuria (50–99/high power field (HPF)), suggestive of deteriorating kidney dysfunction. Cyanotic nephropathy complicated by IgA nephropathy was suspected, prompting his admission for a kidney biopsy.

Upon admission, laboratory findings were as follows: white blood cell count, 7740 / $\mu$ L; red blood cell count, 7.74×10<sup>6</sup> / $\mu$ L; hemoglobin level, 21.3 g/dL; platelet count, 15.0×10<sup>4</sup>/ $\mu$ L; albumin level, 3.8 g/dL; Cre level, 0.78 mg/dL; eGFR, 96.9 mL/min/1.73m<sup>2</sup>; BUN, 11.5 mg/dL; sodium, 140 mEq/L; potassium, 4.5 mEq/L; chloride, 107 mEq/L; IgG, 1376 mg/dL; IgA, 485 mg/dL; IgM, 120 mg/dL; CH50, 42.3 U/mL; C3, 104.8 mg/dL; C4, 18.5 mg/dL; and vascular endothelial growth factor (VEGF), 941

pg/mL. Urinary tests showed a urinary protein/Cre ratio of 2.0 g/gCre and a urinary sediment red blood cell count of 50–99/HPF.

A renal biopsy was conducted, and light microscopy revealed 42 glomeruli, 2 global sclerosis, and 1 focal segmental glomerulosclerosis (FSGS) lesion. The glomeruli displayed hypertrophy, with a mild increase in mesangial cells and matrix. The glomerulus with FSGS exhibited showed a perihilar variant. Immunofluorescence staining showed IgG( $\pm$ ); IgA(+); IgM( $\pm$ ); C1q(-); C3( $\pm$ ); C4(-); CD31(+); CD34(+); and PAL-E(+) (Fig. 1). Electron microscopy showed endothelial cell swelling, subendothelial edema and glomerular basement membrane thickness (777.67 nm) (Fig. 2).

Based on these findings, a diagnosis of CN with endothelial damage was established. Treatment with an SGLT2 inhibitor in addition to an angiotensin-receptor blocker led to a decrease in urinary protein levels from 1.38 g/gCre to 0.28 g/gCre, with stable renal function.

# Discussion

CN is considered a complication of congenital cyanotic heart disease, and the number of cases that develop it increases with prolonged survival. Hongsawong et al. reported that hematocrit>40%; platelets<290,000; and waiting time before surgery are associated with CN [8]. Inatomi et al. [3] also compared patients with CCHD who developed CN with those who had not. Hematocrit values were higher in patients with CN than in those







**Fig. 2** Electron microscopy (EM) showed endothelial cell swelling (original magnification × 4000 (**A**)). EM showed podocyte detachment(original magnification × 1000, (**B**)), subendothelial edema(original magnification × 4000 (**C**)), glomerular basement membrane was thick (777.67 nm, original magnification × 3000, (**D**))

without CN, but no differences were found between these groups with respect to years of disease duration or oxygen saturation. These results support the previously reported hypothesis that increased shear stress induced by polycythemia vera is the main cause of CN; in the glomeruli of patients with CN, glomerular diameter was larger and the number of coagulation hooves was higher than in patients without CN. In the present case, the patient had risk factors relating to hematocrit and platelets, and his histopathology was consistent with CN, as the glomerular diameter was larger and the number of coagulations had increased. In addition, there were no metabolic abnormalities such as diabetes mellitus, but thickening of the basement membrane was observed. The cause of the thickening of the basement membrane has not been mentioned in CN in the past and is not clear, but non-immunological mechanisms may be involved, as there was no deposition of immune complexes.

PAL-E staining is known to primarily react with plasmalemmal vesicle-associated protein-1 (PV-1) expressed in endothelial cells of peritubular capillaries (PTC) and venous endothelial cells, but not with glomerular endothelial cells [8]. However, in transplanted kidneys with extensive glomerular endothelial cell damage, PV-1 was highly expressed in glomerular endothelial cells [7]. Furthermore, PV-1 is co-stained with caveolae. Diabetic nephropathy typically leads to endothelial cell damage, and upregulation of the Cav-1 expression [9, 10]. The aforementioned suggests that endothelial cell damage may be involved in the expression of PV-1. In the present case of CN developed secondary to congenital heart disease, we observed both glomerular hypertrophy and marked glomerular endothelial cell damage, and PAL-E staining, which is typically not seen in glomerular endothelial cells, in the glomerular capillaries. However, the mechanism behind endothelial cell damage in CN has not been previously discussed.

In CN, persistently high levels of VEGF in the bloodstream may contribute to endothelial cell damage. VEGF levels are elevated in diabetic nephropathy and FSGS, and may be associated with renal dysfunction [11]. VEGF stimulates endothelial cells to produce nitric oxide, which, when decreased in bioactivity, can lead to glomerular damage through endothelial cell proliferation and macrophage infiltration. Additionally, VEGF prompts the production of intercellular adhesion molecule-1 (ICAM-1), which may also contribute to endothelial cell damage.

ICAM-1 expression, induced by VEGF, increases vascular permeability by facilitating leukocyte adhesion to vascular endothelial cells [12–14]. In conditions like diabetic nephropathy, increased shear stress triggers ICAM-1 expression, leading to macrophage recruitment



Figure 3 double immunofluorescence microscopy for PAL-E and alpha 2 subtype of collagen type IV (×400, Blue: PAL-E, Red: alpha 2 subtype of type IV collagen, A. Cyanotic nephropathy

[15]. Similarly, reflux nephropathy, which involves shear stress, demonstrates upregulated ICAM-1 expression [16]. This suggests a potential association between ICAM-1 and endothelial cell damage in CN, where shear stress is also a contributing factor. However, the precise relationship remains unclear and necessitates further investigation.

To further clarify the pathogenesis of endothelial cell damage, we performed double staining with alpha2 of collagen type IV and PAL-E to compare with diabetic nephropathy (DN) and transplant glomerulopathy (TGP), which are glomerular diseases causing endothelial cell damage (Fig. 3). DN and TGP showed staining in the glomerular endothelium as in the present case. While a mechanism of endothelial cell damage has been suggested for diabetic nephropathy as described above, Yamamoto et al. [7] reported that endothelial cell damage is associated with PAL-E in TGP. Though endothelial cell damage in TGP has been associated with immunological mechanisms, it is not clear whether CN is caused by immunological or non-immunological mechanisms. It is difficult to conclude whether CN causes endothelial cell damage by immunological or non-immunological mechanisms. In addition, more capillary lumens were stained with PAL-E staining in TGP than in CN and DN in double staining. This may be related to the difference in staining, as CN and DN are associated with shear stress, whereas TGP is associated with immunological mechanisms such as antibodies. However, it was difficult to clarify why segmental endothelial cell damage occurred in this report.

### Conclusion

We encountered a case in which PAL-E indicated potential overexpression of PV-1 in CN, suggesting a link to endothelial cell damage. Despite this observation, many aspects of the pathophysiology of CN remain unclear. Further accumulation of case studies are required for a more comprehensive understanding of its pathogenesis.

### Acknowledgements

Not applicable.

### Authors' contributions

Ushio, Hirata and Manabe wrote the main manuscript text. Suyama, Tanaka, Seki, Kato, Nomura, Nakai, Sumori, Kawaguchi, Kobayashi, Makabe prepared Figs. 1 and 2. Itoh, Taneda and Honda diagnosed pathology. Manabe, Kataoka and Hoshino reviewed the manuscript.

### Funding

This report was not supported by any funding.

### Data availability

No datasets were generated or analysed during the current study.

### Ethics approval and consent to participate

Not applicable for this case report.

### **Consent for publication**

Written informed consent was obtained from the participant for publication of identifying information/images.

### **Competing interests**

The authors declare no competing interests.

Received: 24 May 2024 Accepted: 25 February 2025 Published online: 03 March 2025

### References

- 1. Meessen H, Litton MA. Morphology of the kidney in morbus caeruleus. AMA Arch Pathol. 1953;56(5):480–7.
- Tanaka J, Yasui H, Nakano E, Sese A, Matsui K, Takeda Y, Tokunaga K. Predisposing factors of renal dysfunction following total correction of tetralogy of Fallot in the adult. J Thorac Cardiovasc Surg. 1980;80(1):135–40.
- Inatomi J, Matsuoka K, Fujimaru R, Nakagawa A, Iijima K. Mechanisms of development and progression of cyanotic nephropathy. Pediatr Nephrol. 2006;21(10):1440–5.
- Spear GS. Glomerular alterations in cyanotic congenital heart disease. Bull Johns Hopkins Hosp. 1960;106:347–67.
- Perloff JK, Latta H, Barsotti P. Pathogenesis of the glomerular abnormality in cyanotic congenital heart disease. Am J Cardiol. 2000;86(11):1198–204.
- Hagley MT, Murphy DP, Mullins D, Zarconi J. Decline in creatinine clearance in a patient with glomerulomegaly associated with a congenital cyanotic heart disease. Am J Kidney Dis. 1992;20(2):177–9.
- Yamamoto I, Horita S, Takahashi T, Tanabe K, Fuchinoue S, Teraoka S, Hattori M, Yamaguchi Y. Glomerular expression of plasmalemmal vesicleassociated protein-1 in patients with transplant glomerulopathy. Am J Transplant. 2007;7(8):1954–60.
- Hongsawong N, Khamdee P, Silvilairat S, Chartapisak W. Prevalence and associated factors of renal dysfunction and proteinuria in cyanotic congenital heart disease. Pediatr Nephrol. 2018;33(3):493–501.
- Niemela H, Elima K, Henttinen T, Irjala H, Salmi M, Jalkanen S. Molecular identification of PAL-E, a widely used endothelial-cell marker. Blood. 2005;106(10):3405–9.
- Moriyama T, Tsuruta Y, Shimizu A, Itabashi M, Takei T, Horita S, Uchida K, Nitta K. The significance of caveolae in the glomeruli in glomerular disease. J Clin Pathol. 2011;64(6):504–9.
- Schrijvers BF, Flyvbjerg A, De Vriese AS. The role of vascular endothelial growth factor (VEGF) in renal pathophysiology. Kidney Int. 2004;65(6):2003–17.
- Kurose I, Anderson DC, Miyasaka M, Tamatani T, Paulson JC, Todd RF, Rusche JR, Granger DN. Molecular determinants of reperfusioninduced leukocyte adhesion and vascular protein leakage. Circ Res. 1994;74(2):336–43.
- Del Maschio A, Zanetti A, Corada M, Rival Y, Ruco L, Lampugnani MG, Dejana E. Polymorphonuclear leukocyte adhesion triggers the disorganization of endothelial cell-to-cell adherens junctions. J Cell Biol. 1996;135(2):497–510.
- Bolton SJ, Anthony DC, Perry VH. Loss of the tight junction proteins occludin and zonula occludens-1 from cerebral vascular endothelium during neutrophil-induced blood-brain barrier breakdown in vivo. Neuroscience. 1998;86(4):1245–57.
- Chiu JJ, Lee PL, Chen CN, Lee CI, Chang SF, Chen LJ, Lien SC, Ko YC, Usami S, Chien S. Shear stress increases ICAM-1 and decreases VCAM-1 and E-selectin expressions induced by tumor necrosis factor-[alpha] in endothelial cells. Arterioscler Thromb Vasc Biol. 2004;24(1):73–9.
- Unemoto K, Chertin B, Shima H, Bi X, Sakai M, Puri P. ICAM-1 expression is upregulated in reflux nephropathy. Pediatr Surg Int. 2003;19(5):391–4.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.